NEW AROMATIC SESQUITERPENOIDS FROM THE RED ALGA LAURENCIA OKAMURAI YAMADA (1)

Minoru Suzuki and Etsuro Kurosawa Department of Chemistry, Faculty of Science, Hokkaido University, Sapporo, Japan

(Received in Japan 29 March 1978; received in UK for publication 15 May 1978)

Many halogenated metabolites have been isolated from the marine red algae genus <u>Laurencia</u> (Rhodomelaceae), and various species of <u>Laurencia</u> are known to contain, although some overlap is noted, unique halogenated compounds (species-specific ?)(2).

In connection with our interest in algal natural products we have examined the essential oils of the red algae <u>L</u>. <u>okamurai</u> Yamada collected from two different locations. Previously, we reported (3) the isolation of laurinterol (<u>1a</u>), debromolaurinterol (<u>1b</u>), aplysin (<u>5a</u>) and debromoaplysin (<u>5b</u>) as the major components and aplysinol (<u>6</u>) as the minor one, from <u>L</u>. <u>okamurai</u> collected in August at Hakata-shima, the Inland Sea of Japan. We collected <u>L</u>. <u>okamurai</u> early in June, 1977, at Nyudogatane, Okino-shima, Tosa Province, Japan, and examined. This alga displayed no aplysin, debromoaplysin and aplysinol, but instead yielded three new sesquiterpenes and several known sesquiterpenes, which have previously been described from other <u>Laurencia</u> species, together with laurinterol and debromolaurinterol. Reported below are the structures of these interesting new compounds.

Half-dried algae (ca. 700g) were extracted with methanol, and the methanol extract (ca. 10g), after separation of the acidic and basic components, was fractionated by column chromatography on neutral alumina; (i) hexane fraction, (ii) hexane-benzene fraction and (iii) benzene fraction.

Sesquiterpene phenols. Benzene fraction (iii) gave laurinterol (1a) (30% of neutral oil) and debromolaurinterol (1b)(20%) along with isolaurinterol (2a)(0.5%) and debromoisolaurinterol (2b)(0.5%) (4). Hexane-benzene fraction (ii) was rechromatographed on silica gel. Benzene eluate consisted of a complex mixture of fatty acid methyl ester. Hexane eluate gave new halogenated phenol (3)(0.1%), designated as neolaurinterol, $C_{15}H_{19}OBr$ (m/e 296 and 294, M⁺); m.p. 62-63°; [α]_D-14° (c 0.57; CHCl₃); $\nu_{max}(CHCl_3)$ 3580, 1607, 1190 and 1135 cm⁻¹; δ (100 MHz, CCl₄) ca. 0.5 (2H, m), ca. 1.1 (1H, m), 1.27, 1.37, 2.33 (each 3H, s), 5.64 (1H, s; D₂0 exchangeable), 6.61 (1H, d, J=8 Hz) and 7.28 (1H, d, J=8 Hz). The pmr spectrum of neolaurinterol (3) displayed the signals comparable to those of laurinterol (1a) except for in the lower magnetic field region (aromatic protons of la; each singlet at δ 6.42 and 7.47).

$$X$$
OH
OH
 S
O

Treatment of $\underline{3}$ with acetic anhydride and pyridine at room temperature (overnight) afforded an acetate $(\underline{4})$ in low yield (40%), whereas $\underline{1a}$ was acetylated quantitatively under the same condition. The above-mentioned data indicates that the $\underline{\text{ortho}}$ -position of the hydroxyl group of $\underline{3}$ might be substituted by the bromine atom. Confirmation of the structure of $\underline{3}$ was obtained by the following reaction. Debromolaurinterol $(\underline{1b})$ was treated with NBS in CCl₄ (5) at room temperature (1 hr) to yield a brominated product, $[\alpha]_{\overline{0}}$ -17° (c 1.19), (70% yield), which was identical with natural $\underline{3}$ by the comparison of spectral data. Thus, neolaurinterol is represented by formula $\underline{3}$, including absolute stereochemistry.

Sesquiterpene ethers. Hexane fraction (i) was rechromatographed on silica gel. Earlier hexane eluates consisted of a mixture of hydrocarbons (vide infra) and successive eluates were further submitted to thin layer chromatography over silica gel to yield two new ether compounds. The less polar halogenated compound (7) (0.2%), designated as isoaplysin, $C_{15}H_{19}OBr$ (m/e 296 and 294, M⁺), $[\alpha]_D$ -33° (c 0.69), had the following spectral characteristics; ν_{max} (film) 1620, 1597, 1501, 1275, 1265, 950 and 802 cm⁻¹; δ 1.09 (3H, d, J=6 Hz), 1.50 (3H, s), 2.26 (3H, s), 3.55 (2H, s), 6.50 (1H, br. s), 6.58 (1H, br. d, J=8 Hz) and 6.82 (1H, d, J=8 Hz); m/e (relative intensity) 296, 294 (47), 281, 279 (7), 239, 237 (7), 215 (35), 201 (20), 199 (12), 173 (16), 159 (100), 145 (15), 135 (75), 121 (31), 115 (11), 107 (11) and 91 (12).

 $\frac{\text{TABLE}}{\text{C nmr shifts (δ, ppm downfield from (CH}_{3})_{4}\text{Si, CDCl}_{3})}$

Compound Carbon	<u>5b</u>	7	<u>9a</u>	<u>9b</u>	9c	10
	158.8(s)	158.7(s)	152.3(s)	151.6(s)	152.3(s)	152.4(s)
	137.7(s)	138.2(s)	136.4(s)	136.5(s)	137.4(s)	137.0(s)
	133.3(s)	132.9(s)	130.1(s)	130.3(s)	127.7(s)	130.5(s)
	122.5(d)	122.1(d)	128.4(d)	128.6(d)	124.7(d)	124.0(d)
	120.6(d)	121.4(d)	117.4(d)	117.5(d)	121.2(d)	120.6(d)
	109.3(d)	109.3(d)	114.4(s)	114.9(s)	115.8(d)	115.8(d)
C-2	98.8(s)	97.1(s)	46.4(d)	43.1(d)	44.3(d)	41.0(s)
C-1	54.0(s)	55.5(s)	44.9(s)	44.9(s)	45.1(s)	45.7(s)
C-3	46.1(d)	43.7(d)	85.3(s)	87.4(s)	85.7(s)	86.2(d)
C - 5	42.6(t)	42.6(t)	42.2(t)	41.8(t)	41.8(t)	41.4(t)
C-4	31.2(t)	31.5(t)	37.3(t)	33.0(t)	34.5(t)	30.1(t)
	23.5(q)	34.5(t)	23.0(q)	66.0(t)	36.4(t)	21.0(q)
	21.4(q)	22.9(q)	22.5(q)	22.5(q)	21.0(q)	20.5(q)
	20.0(q)	21.5(q)	20.4(q)	20.0(q)	20.5(q)	18.2(q)
	13.1(q)	13.8(q)	7.4(q)	7.9(q)	7.2(q)	15.0(q)

The pmr and cmr (table) spectra of 7 are very similar to those of debromoaplysin (5b). In the pmr spectra of 7 and 8, can be written for isoaplysin.

The choice in favour of structure 7 for isoaplysin was made with the aid of 13 C chemical shift. In the cmr spectra of 7 and 5b, the distinct difference, 2.4 ppm, was observed for the signals of C-3 appeared at 43.7 in 7 and 46.1 in 5b. The comparative upfield shifts were also observed for the signals of C-2 and C-4 in the cmr spectra of filiformin (9a) (6), filiforminol (9b) (6) and 9c (7). These upfield shifts were explained by the effects due to additional γ -substituents, and, consequently, the BrCH₂- group in 7 must be attached to C-2. The structure 7 for isoaplysin will also be explicable well from the biogenetic consideration, demonstrating that the presence of substituents has not been recognized on the benzylic methyl group at C-1 of this type of sesquiterpenes.

The polar nonhalogenated compound $(\underline{10})(0.4\%)$, $C_{15}H_{20}O$ (m/e 216, M⁺), $[\alpha]_D$ -25° (c 0.68), exhibited in its spectra [v_{max} (film) 1620, 1578, 1505, 1397, 1379, 1247, 1160, 1039 and 808 cm⁻¹; δ 0.87, 0.96, 1.22, 2.20 (each 3H, s), 4.02 (1H, br. d), 6.40 (1H, br. s), 6.48 (1H, br. d, J=8 Hz) and 6.85 (1H, d, J=8 Hz); m/e 216 (100), 201 (75), 173 (71), 159 (40), 148 (80), 147 (30), 145 (20), 135 (22), 133 (18), 121 (17), 115 (11), 105 (10) and 91 (15)] the presence of three tertiary methyls, including gem-dimethyl group (cmr (table) 41.0 (s)), one of which was shifted to higher magnetic field by an anisotropic effect of the aromatic nucleus, and aromatic methyl and 2,5-disubstituted phenol ether group. In addition, the cmr spectrum of 10 indicates the presence of the aromatic ring and further no double bond, and, hence, the compound (10), having six degrees of unsaturation, must contain other two rings, one of which is the ether ring and the other may be a five-membered carbocyclic ring. Furthermore, in the cmr spectrum of 10, the peaks at 86.2 (d) and 45.7 (s), which are assignable to C-3 and C-1, respectively, are similar to those of (9a) (6) and (9b) (6) and 9c (7). This indicates that the ether oxygen is combined with C-3, having one proton. In view of the above-mentioned data and the standpoint of biogenesis, the structure of this ether compound would be represented as 10.

Sesquiterpene hydrocarbons. Repeated silica gel column and thin layer chromatography of the mixture of hydrocarbons yielded isolaurene (0.7%), α -bromocuparene (1%) and α -bromoisocuparene (0.3%), previously isolated from \underline{L} . glandulifera Kützing and L. nipponica Yamada (8)(9).

References

- (1) Part XXIX of "Constituents of Marine Plants". Part XXVIII, T. Suzuki, A. Furusaki, N. Hashiba and E. Kurosawa, Tetrahedron Letters, 3731 (1977).
- (2) (a) W. Fenical, J. Phyco., 11, 245 (1975); (b) D. J. Faulkner, Tetrahedron, 33, 1421 (1977); (c) S. J. Wratten and D. J. Faulkner, J. Org. Chem., 42, 3343 (1977); (d) J. W. Blunt, M. P. Hartshorn, T. J. McLennan, M. H. G. Munro, W. T. Robinson and S. C. Yorke, Tetrahedron Letters, 69 (1978).
- (3) T. Irie, M. Suzuki and Y. Hayakawa, <u>Bull. Chem. Soc. Japan</u>, <u>42</u>, 843 (1969).
- (4) T. Irie, M. Suzuki, E. Kurosawa and T. Masamune, Tetrahedron, 26, 3271 (1970)
- (5) D. E. Pearson, R. D. Wysong and C. V. Breder, J. Org. Chem., 32, 2358 (1967).
- (6) R. Kazlauskas, P. T. Murphy, R. J. Quinn and R. J. Wells, <u>Aust. J. Chem.</u>, 29, 2533 (1976).
- (7) M. Suzuki and E. Kurosawa, Tetrahedron Letters, 4817 (1976).
- (8) T. Irie, T. Suzuki, Y. Yasunari, E. Kurosawa and T. Masamune, <u>Tetrahedron</u>, 25, 459 (1969).
- (9) T. Suzuki, M. Suzuki and E. Kurosawa, Tetrahedron Letters, 3057 (1975).